Functional characterization of VvCAX3: a grapevine cation/H⁺ exchanger that transports Ca²⁺ and other cations

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INTRODUCTION

Grapevines are economically one of the most important fruit species worldwide. Thus, it is essential for winegrowers to guarantee fruit quality upon adverse climate conditions, including heavy rains before harvest that cause severe skin cracking and fruit spoilage. Calcium (Ca²⁺) is beneficial to the fruit integrity, and thus for quality, due to its key structural and signaling roles, acting as osmoticum within vacuoles, as strengthening agent in cell walls, and as secondary messenger for a large number of abiotic stress responses [1]. In fact, a close relationship has been demonstrated between increased tomato fruit integrity, increased Ca²⁺ levels and increased activity of CAX-type cation/H⁺ exchangers (CAXs), that appear to predominantly reside on the vacuole [2]. Therefore, the identification and characterization of grapevine CAX transporters is a landmark towards understanding calcium dynamics in the grape berry.



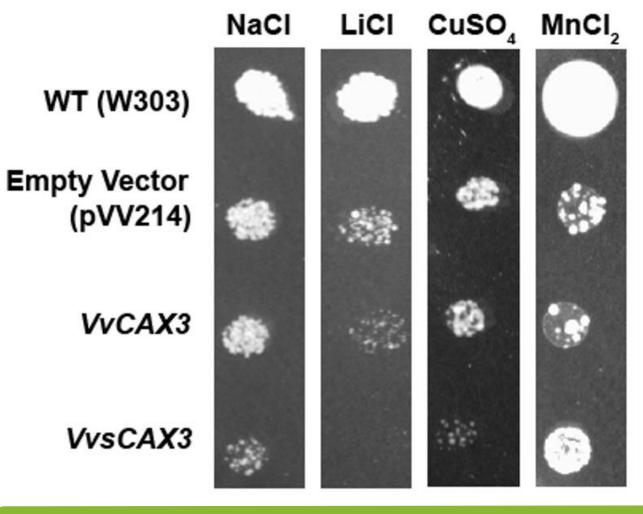
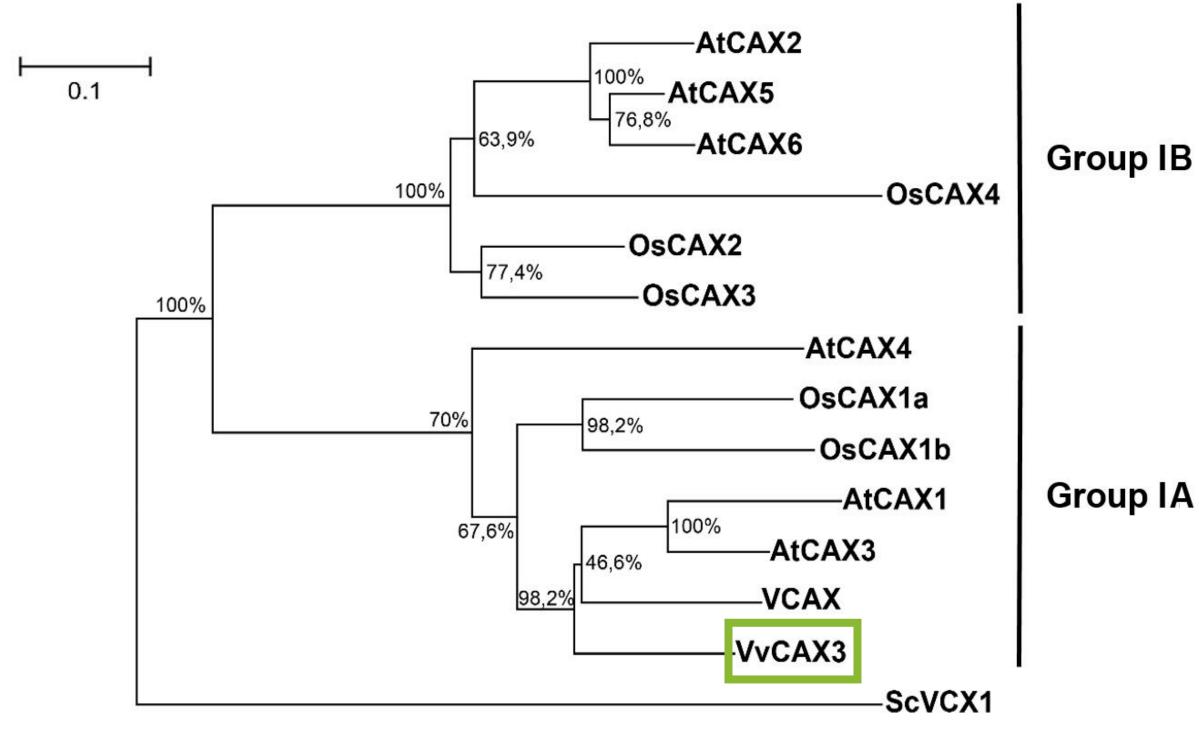


Fig. 4 – Growth assays of *S. cerevisiae* K667 strain carrying *VvCAX3*, the truncated *VvsCAX3*, or the empty vector. Wild-type strain (WT; W303) was used as positive control. Cells were platted as drops in YPD medium supplemented with NaCl (500 mM), LiCl (50 mM), CuSO₄ (6 mM) and MnCl₂ (750 μ M), for 2 days.

RESULTS

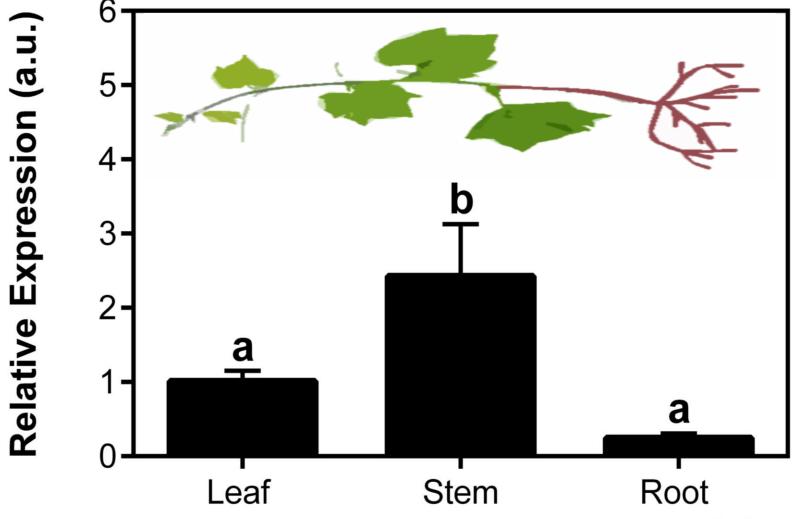
Phylogenetic and topologic analysis of VvCAX3

Prospection of the grapevine genome allowed the identification of *VvCAX3*, a putative calcium-H⁺ antiporter belonging to the family of cation-H⁺ exchangers (**Fig. 1**), with a well defined topological structure (**Fig. 2**).



Expression of VvCAX3 in grapevine

To assess the involvement of VvCAX3 in grapevine calcium dynamics, its expression was studied by real-time PCR in different plant organs, berry developmental stages and upon elicitation with specific compounds.



The highest expression of *VvCAX3* was detected in the grapevine **stems** followed by the leaves and the roots (**Fig. 5**).

Fig.5 - Expression of *VvCAX3* by real-time PCR in grapevine organs cv. "Trincadeira".

Transcript levels were higher in the **green stage** of berry development, followed by veraison and mature stages (**Fig. 6**), consistent with the pattern of calcium accumulation in fruits [4].

Fig.6 - Expression of *VvCAX3* throughout grape berry development cv. "Vinhão", by

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Fig.1 – Phylogenetic relationship between VvCAX3 and other CAX proteins from *Arabidopis thaliana* (AtCAX1, AtCAX2, AtCAX3, AtCAX4, AtCAX5, AtCAX6), *Oryza sativa* (Os CAX1a, OsCAX1b, OsCAX2, OsCAX3, OsCAX4), *Saccharomyces cerevisiae* (ScVCX1) and *Vigna radiata* (VCAX).

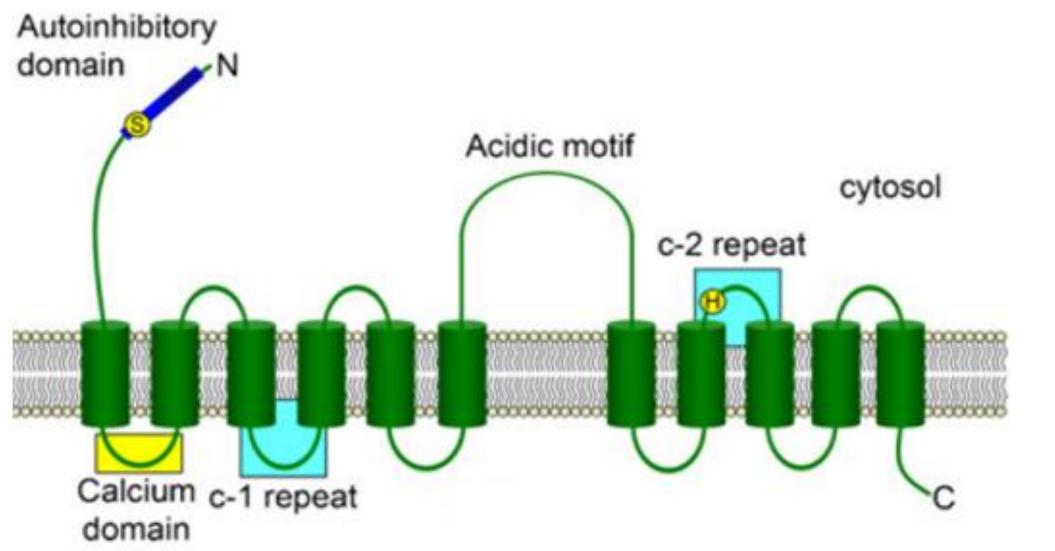


Fig.2 – Proposed topological model of plant CAX proteins (Adapted from [3]).

Considering the predicted topological model, two forms of *VvCAX3* were cloned: the whole gene (*VvCAX3*) and a truncated form without the codons encoding the **autoinhibitory domain** (*VvsCAX3*).

*Functional complementation of a yeast mutant for Ca*²⁺ *transport* The functional characterization of VvCAX3 was performed following heterologous expression on a yeast strain characterized for its high sensitivity to Ca²⁺ (**Fig. 3**, **Fig. 4**). real-time PCR.



VvCAX3 expression increased in grape cell suspensions (CSB) upon elicitation with Ca²⁺ (100 mM), Na⁺ (100 mM) and methyl jasmonate (Meja, 20 μ M) (**Fig. 7**). In contrast, Mn²⁺ (5 mM) and sucrose (150mM) caused an apparent reduction in transcript levels.

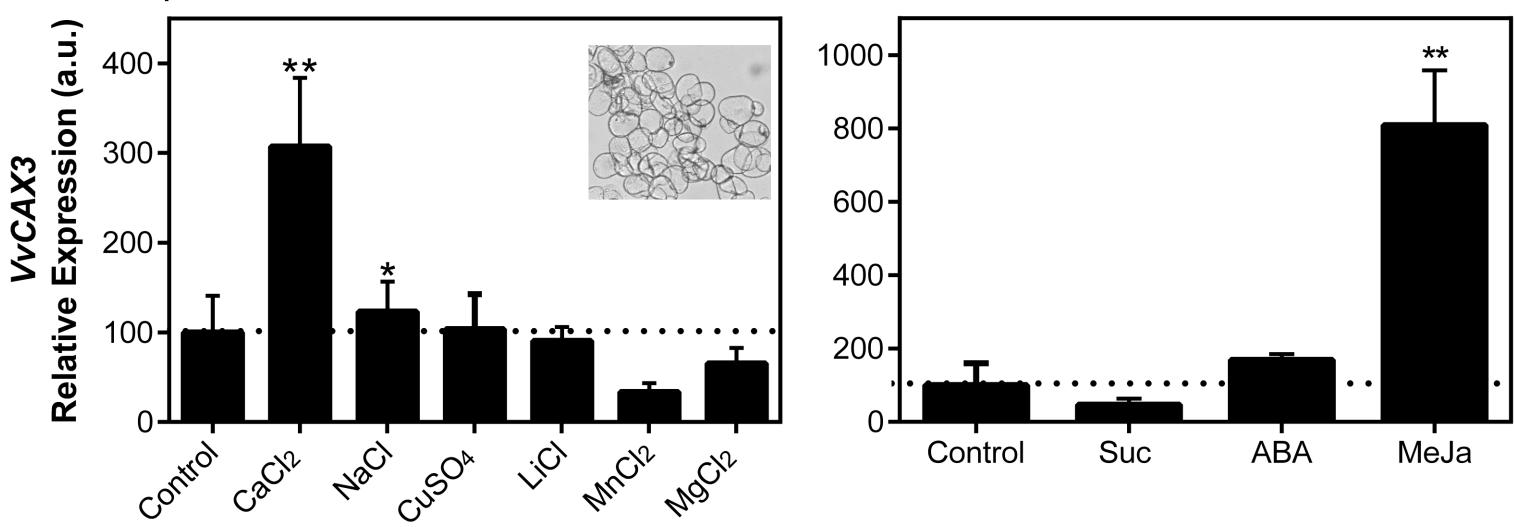


Fig.7- Expression of *VvCAX3* in grape cell suspensions (CSB – Cabernet Sauvignon Berry), by real-time PCR. Cells were elicitated for 12 hours with: 100 mM CaCl₂, 100 mM NaCl, 150 μ M CuCl₂, 75 mM LiCl, 5 mM MnCl₂, 100 mM MgCl₂, 150 mM sucrose, 20 μ M abscisic acid (ABA) and 20 μ M methyl jasmonate (MeJa).

CONCLUDING REMARKS

 VvsCAX3 lacking the CAX autoinhibitory domain was able to restore the growth defect of the yeast strain K667 at high Ca²⁺ levels, validating the role of the protein in Ca²⁺ transport.

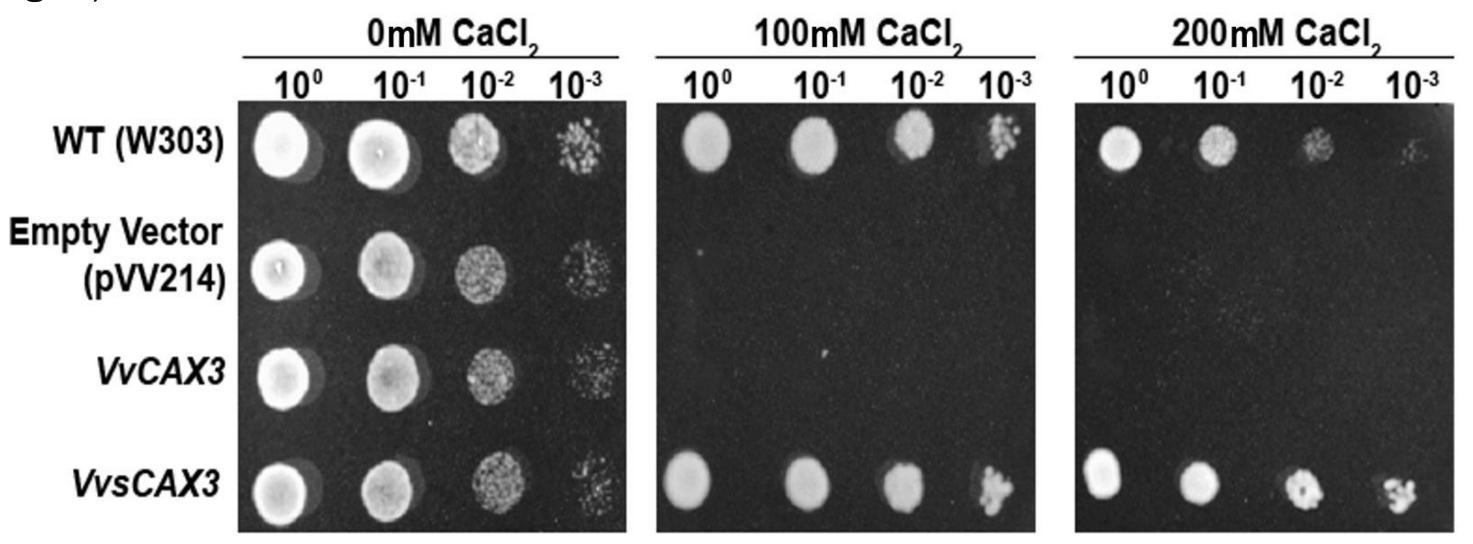


Fig.3 – Complementation assays of *S. cerevisiae* K667 strain by *VvCAX3* and the truncated *VvsCAX3*. Cells were transformed with the vector pVV214 alone (empty vector) or with the same vector carrying *VvCAX3* or *VvsCAX3*. Wild-type strain (WT; W303) was used as positive control. Several fold dilutions were platted as drops in YPD medium supplemented with CaCl₂ (0, 100, 200 mM), and growth was analyzed after 2 days.

- Likewise, VvsCAX3 restored the growth defect of the yeast strain at high Mn²⁺ levels, but increased its sensitivity for Na⁺, Li⁺ and Cu²⁺, suggesting its additional involvement in the transport of these cations.
- VvCAX3 transcripts were detected in all grapevine organs, and expression decreased gradually during grape berry development, in accordance to the pattern of calcium accumulation in the fruit.
- VvCAX3 expression was upregulated by Ca²⁺ and Na⁺, further supporting its involvement in the homeostasis of calcium and other cations in grapevine.

Acknowledgements: VM is supported by FCT-Portuguese Foundation for Science and Technology (post-doctoral grant SFRH/BPD/107905/2015). This work was supported by European Union Funds (FEDER/COMPETE Operational Competitiveness Programme) and Portuguese national Funds (FCT-Portuguese Foundation for Science and Technology): KBBE-2012-6-3117 "Inovinne", FCOMP-01-0124-FEDER-022692 and PTDC/AGR-ALI/100636/2008. This work also benefited from the networking activities within the European COST ACTION (FA1106 'QualityFruit').

References

[1] Dodd, A.N., Kudla, J. & Sanders, D., 2010. The language of calcium signaling. Annual Review of Plant Biology, 61(1), pp.593–620.
[2] Martinoia, E., Maeshima, M. & Neuhaus, H.E., 2007. Vacuolar transporters and their essential role in plant metabolism. Journal of Experimental Botany, 58(1), pp.83–102.

[3] Pittman, J.K., 2011. Vacuolar Ca²⁺ uptake. Cell Calcium, 50(2), pp.139–146

[4] Martins V, Cunha A, Gerós H, Hanana M, Blumwald E (2012) Mineral compounds in grape berry. In: The Biochemistry of the Grape Berry, Gerós H, Chaves M-M, Delrot S (eds), Bentham Science Publishers, pp 23-43.