

SURFACE MODIFICATION OF BACTERIAL CELLULOSE BY PLASMA TREATMENTS

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INTRODUCTION

Plasma technique is a convenient method to introduce functional groups or chains onto the materials' surface, including those with complex shapes; being conducted under vacuum, the treatment is pervasive, an advantage in the case of scaffolds with interpenetrating porous structures, often used for tissue engineering purposes [1].

Bacterial cellulose (BC) is a biocompatible material with unique properties that makes it an attractive material for biomedical applications. With the major goal of improving the biocompatibility of BC, and in line with previous research within our group [2], this work aimed at assessing the effect of plasma treatments on the in vitro interaction of BC with animal cell lines.

MATERIALS AND METHODS

BC production

- Cultivation of Gluconacetobacter xylinum (ATCC 53582) in static culture, at 30 $^{\circ}$ C in Hestrin-Shramm (HS) medium [3].

• Purification BC pellicles in a 4% (w/v) NaOH at 50 °C, overnight.

Freeze-drying of BC pellicles.

Plasma treatments

Ar flow (80 sccm) and

N₂ flow (40 sccm), current (0.75 A), time (1200 s);
N₂ flow (40 sccm), current (1 A), time (700 s);
N₂ flow (35 sccm), current (0.4 A), time (1800 s);
N₂ flow (50 sccm), current (0.5 A), time (1800 s);
N₂ flow (35 sccm), current (0.5 A), time (300 s);
N₂ flow (55 sccm), current (0.5 A), time (1200 s);
N₂ flow (5 sccm), current (0.5 A), time (1200 s);
N₂ flow (80 sccm), current (0.5 A), time (1200 s);
Ar flow (80 sccm), current (0.5 A), time (600 s).

• Ar flow (80 sccm), current (0.5 A), time (60

Surface analysis

X-ray photoelectron spectroscopy (XPS);

• Scanning electron microscopy (SEM).

Cell adhesion and proliferation assays

Preformed using Human Microvascular Endothelial Cells (HMEC-1) as described by Pertile and co-workers [2].

RESULTS

X-ray photoelectron spectroscopy (XPS)

 $\begin{array}{l} \textbf{Table 1} \textbf{-} BC \ \text{and} \ BCP \ \text{elemental composition analyzed by XPS. The BC} \\ \text{samples were treated using the following conditions: 1) 80 sccm Ar e 40 sccm} \\ N_2, \ 0,75 \ A, \ 1200 \ s; \ 2) \ 80 \ \text{sccm Ar} \ e \ 40 \ \text{sccm} \ N_2, \ 1 \ A, \ 700 \ s; \ 3) \ 80 \ \text{sccm Ar}, \ 35 \ \text{sccm} \ N_2, \ 0,4 \ A, \ 1800 \ s. \end{array}$

Sample Oxygen (%) Carbon (%) Nitrogen (%)

1	32.69	64.12	3.19
2	34.90	63.16	1.94
3	28.05	68.18	3.77
4	35.58	59.84	4.58

From table 1 a longer plasma treatment allow for a higher N_2 incorporation.

The N_2 content values of samples 3 and 4 shows that the pellicle which was treated with a higher N_2 flow had more nitrogen incorporation.

Scanning electron microscopy (SEM)

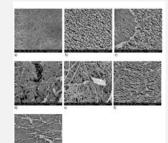


Figure 1 - SEM micrographs of untreated BC a) and treated BC b) to g). The treatment conditions were the following: b) 80 sccm Ar, 35 sccm N₂, 0,4 A, 1800 s; c) 80 sccm Ar, 50 sccm N₂, 0,4 A, 1800 s; d) 80 sccm Ar, 35 sccm N₂, 0.5 A, 600 s; e) 80 sccm Ar, 35 sccm N₂, 0.5 A, 300 s; f) 80 sccm Ar, 5 sccm N₂, 0.5 A, 1200 s; g) 80 sccm Ar, 5 sccm N₂, 0.5 A, 600 s.

The treatment conditions lead to significant surface morphology change resulting in the destruction of the BC fibres and in the formation of a heterogeneous surface when higher N_2 flows were used as can be evidenced by the image 1c).

Cell adhesion and proliferation assays

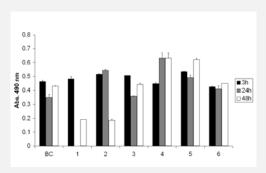


Figure 2 - MTS assays of HMEC-1 cultured on untreated BC and BC treated with the following conditions: 1 - 80 sccm Ar, 0.5 A; 1200 s; 2 - 80 sccm Ar, 0.5 A, 600 s; 3 - 80 sccm Ar, 35 sccm N₂, 0.5 A, 600 s; 4 - 80 sccm Ar, 35 sccm N₂, 0.5 A, 300 s; 5 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 1200 s; 6 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 4 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 1200 s; 6 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 1200 s; 6 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 4 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 5 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 1200 s; 6 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 6 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 6 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 6 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 6 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 6 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 6 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 6 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 6 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 6 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 6 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 6 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 6 - 80 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 6 - 80 sccm Ar, 5 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 6 - 80 sccm Ar, 5 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 6 - 80 sccm Ar, 5 sccm Ar, 5 sccm N₂, 0.5 A, 600 s; 6 - 80 sccm Ar, 5 s

Ar plasma treated BC cause cell death after 48 h (treatments 1 and 2), it should be mentioned that this effect did not present changes for 600s (treatment 1) or 1200s (treatment 2) treatment times. The effect above described is reversed by the presence of N_2 on the treatment gas (3 trough 6 treatments).

CONCLUSIONS

The N_2 incorporation is treatment time and N_2 flow dependent; The plasma treatments induced morphological changes on the BC surface;

Harsher treatments resulted in the formation of heterogeneous surfaces;

BC treated with Ar plasma induced cell death and that this effect was reversed by the presence of N_2 on the treatment gas.

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