**In vitro Antifungal Effect of EDTA Disodium Salt in Tested Black Aspergilli**

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**Abstract:** The antifungal effect of Na$_2$EDTA on *Aspergillus carbonarius*, *A. tibericus*, an ochratoxigenic *A. niger* and a non-ochratoxigenic *A. niger* strain was studied. Also, the effect of Na$_2$EDTA on the production of ochratoxin A by *A. carbonarius* and the ochratoxigenic *A. niger* was evaluated. The poisoned food technique was used with CYA medium supplemented with 0, 1 and 10 mmol L$^{-1}$ of Na$_2$EDTA. The colony diameters were recorded daily and the amount of ochratoxin A produced was quantified every two days. Significant reductions of growth rates were observed in the presence of Na$_2$EDTA being the calculated EC$_{50}$ of 2.1 mmol L$^{-1}$ for *A. carbonarius*, 0.9 mmol L$^{-1}$ for *A. tibericus*, 2.0 mmol L$^{-1}$ for the ochratoxigenic *A. niger* and 4.1 mmol L$^{-1}$ for the non-ochratoxigenic *A. niger*. Furthermore, 10 mmol L$^{-1}$ Na$_2$EDTA delayed the production of ochratoxin A and reduced the levels in approximately 99% during 8 days. Na$_2$EDTA is frequently used in the food industry and in agriculture agrochemicals and its effects on ochratoxigenic black aspergilli is not well known. This study showed that Na$_2$EDTA can significantly reduce the growth rates of tested fungi and its ochratoxin A production.

**Key words:** *Aspergillus carbonarius*, *Aspergillus tibericus*, *Aspergillus niger*, ochratoxin A, EDTA, fungicide

**INTRODUCTION**

Ochratoxin A (OTA) is a mycotoxin produced by some *Aspergillus* and *Penicillium* species in several agricultural commodities. Therefore, OTA is found in food products such as breakfast cereals, coffee, cocoa products, dried vine fruits, dried figs, beer and wine. The presence of OTA in grapes is due to *A. carbonarius* and to some ochratoxigenic isolates of the *A. niger* aggregate (Serra et al., 2003; Gomez et al., 2006). *A. carbonarius* isolates are producers of higher amounts of OTA and are more frequent on grapes than the ochratoxigenic isolates of the *A. niger* aggregate (Serra et al., 2006; Esteban et al., 2004). *Aspergillus niger* aggregate strains are also able to degrade OTA into the less toxic ochratoxin a (Vanga et al., 2000; Abrunhosa et al., 2002).

Ethyleneaminoethanetricarboxylic Acid (EDTA) is a strong chelating agent that forms several metal and salt complexes. EDTA and EDTA-complexes are widely used in cleaners and detergents, in agriculture or in food processing (Oviédo and Rodriguez, 2003). In the food industry, it is mostly used to remove metallic tastes by sequestering metal ions released during the processing or storage (e.g., in canned beans). In USA, Na$_2$EDTA concentrations of 36 to 500 ppm, which corresponds to 0.1-1.5 mmol L$^{-1}$ in solution, are allowed in some food products (Heimbach et al., 2000). In agriculture, EDTA is used in agrochemical products to stabilize formulations or to provide micronutrients such as zinc, manganese, iron, copper, magnesium, calcium and potassium. EDTA is also recognized as an
antibacterial agent which disrupts the membrane integrity and as a potentiator of other lethal agents (Oita, 2003). EDTA antifungal properties were mainly tested on yeasts (Siqueira and Sen, 2004; Kubo et al., 2005), being nevertheless reported its synergetic effect with another antifungal agent on the reduction of pulmonary aspergillosis (Hachem et al., 2006). Its effect in controlling mildew on tomato leaves was also reported (Ehrat et al., 2002).

In this study, the in vitro effect of Na<sub>2</sub>EDTA on the growth of an A. carbonarius, an A. ibericus, an ochratoxigenic A. niger and a non ochratoxigenic A. niger strains was assessed. All these strains were isolated from grapes.

MATERIALS AND METHODS

Chemical Material

Ethylendiaminetetraacetic acid disodium salt (Na<sub>2</sub>EDTA) from Merck commercialized as Titriplex III.

Biological Material

Ochratoxigenic Aspergillus carbonarius strain MUM 03.59, non-ochratoxigenic A. ibericus strain MUM 03.49, ochratoxigenic A. niger strain MUM 03.57 and non-ochratoxigenic A. niger strain MUM 03.58.

Growth Conditions in Presence of Na<sub>2</sub>EDTA

The antifungal activity of Na<sub>2</sub>EDTA was evaluated by the agar dilution method using Czapek Yeast Extract Agar medium (CYA) (Mares et al., 2004). The medium was supplemented with 0, 1 or 10 mmol L<sup>-1</sup> of Na<sub>2</sub>EDTA (CYA, CYA+1 m and CYA+10 m, respectively). Strains were first grown in plates with MEA medium (Blakeslee formula), for 7 days, at 25°C, in the dark, for inoculum generation. For each strain, a spore suspension with 2.5×10<sup>6</sup> spores mL<sup>-1</sup> was prepared in semi solid agar (0.2% agar and 0.05% Tween 80) using a NeuBauer chamber (Aberkane et al., 2002). Each strain was centrally inoculated in triplicate in each media with 10 μL of the respective spore suspension and incubated at 25°C, in the dark. The colony diameters were recorded daily and growth rates were calculated by linear regression of colony diameters against days. The concentration of Na<sub>2</sub>EDTA at which survival was 50% (EC<sub>50</sub>) was determined by fitting the experimental data to a four-parameter logistic model (Hill equation) using computer curve-fitting software (Prism 4, GraphPad Software, Inc, San Diego, CA, USA).

Ochratoxin A Analysis

Every two days, plates were extracted with methanol to quantify the amount of OTA produced. A modification of the method presented by Bragulat et al. (2001) was used as follows. The mycelia and media were cut and transferred to tubes with 20 mL of methanol. The tubes were vigorously vortexed and allowed to extract overnight. Methanol extracts were filtered with a 0.45 μm syringe filter of PTFE (Tefnomokroma) and 1 mL of the filtrate dried at 50°C with a gentle stream of nitrogen in a clean vial. Dried residues were resuspended in 1 mL of HPLC mobile phase and analyzed by high-performance liquid chromatography. The HPLC apparatus consisted of a Varian 9002 pump equipped with a Jasco FP-920 fluorescence detector (λ<sub>ex</sub> = 333 nm, λ<sub>em</sub> = 460 nm) and a Marathon Basic autosampler. The analytical column was a C<sub>18</sub> reversed-phase YMC-Pack ODS-AQ (250×4.6 mm and 5 μm) fitted with a precolumn with the same stationary phase. The mobile phase was a mixture of acetonitrile/water/acetic acid (99/99/2, v/v/v) filtered and degassed. The flow rate was set to 0.8 mL min<sup>-1</sup> and the column temperature to 30°C. The loop volume was 100 μL. Calibration curves were prepared with standards of OTA (Sigma).
Statistical Analysis

All statistical analyses were performed with the Statistic Package for Social Sciences (SPSS) version 15.0. Means were compared by analysis of variance followed by Duncan’s post-test being the differences considered statistically significant when \( p<0.05 \).

RESULTS AND DISCUSSION

The tested strains were found to be susceptible to the presence of Na₂EDTA in culture media. Na₂EDTA produced a significant increase in the colonies lag phase (Fig. 1) and a significant decrease on colonies growth rates (Table 1). In the presence of 1 mmol L⁻¹ of Na₂EDTA, the growth rate of A. carbonarius, A. ibericus, A. niger MUM 03.57 and A. niger MUM 03.58 was reduced in 32, 56, 36 and 16%, respectively. When 10 mmol L⁻¹ of Na₂EDTA was used, reductions in growth rate of 88, 89, 87 and 82% were obtained, respectively. The concentration of Na₂EDTA which reduces growth

![Graphs showing radial growth of different strains](image)

Fig. 1: Daily radial growth of A) A. carbonarius MUM 03.59, B) A. ibericus MUM 03.49, C) A. niger MUM 03.57 and D) A. niger MUM 03.58 cultivated in: (- - -) CYA, (• - •) CYA+1 m and (-Δ-) CYA+10 m. Values presented are the mean of three replicates.

<table>
<thead>
<tr>
<th>Medium</th>
<th>A. carbonarius</th>
<th>A. ibericus</th>
<th>A. niger</th>
<th>A. niger</th>
</tr>
</thead>
<tbody>
<tr>
<td>CYA</td>
<td>1.65±0.02b</td>
<td>1.88±0.01c</td>
<td>1.72±0.01b</td>
<td>1.38±0.01c</td>
</tr>
<tr>
<td>CYA+1 m</td>
<td>1.25±0.01c</td>
<td>0.93±0.01</td>
<td>1.18±0.01</td>
<td>1.27±0.01</td>
</tr>
<tr>
<td>CYA+10 m</td>
<td>0.15±0.02</td>
<td>0.15±0.01</td>
<td>0.26±0.02</td>
<td>0.24±0.02</td>
</tr>
</tbody>
</table>

Values are the mean of three replicates±Standard Deviation (SD). Data marked with different letter(s) in each column are significantly different at \( p<0.05 \) for the Duncan’s post hoc test.
Table 2: The 50% effective concentration (EC₅₀) of Na₂EDTA for the different black aspergilli tested

<table>
<thead>
<tr>
<th>Strains</th>
<th>EC₅₀ (µM)</th>
<th>95% CI</th>
<th>p² (r²)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>A. carbonarius</em> (MUM 03.59)</td>
<td>2.1</td>
<td>1.59 to 2.23</td>
<td>0.9993</td>
</tr>
<tr>
<td><em>A. ibericus</em> (MUM 03.49)</td>
<td>0.9</td>
<td>0.92 to 0.95</td>
<td>0.9999</td>
</tr>
<tr>
<td><em>A. niger</em> (MUM 03.57)</td>
<td>2.0</td>
<td>1.58 to 2.10</td>
<td>0.9998</td>
</tr>
<tr>
<td><em>A. niger</em> (MUM 03.58)</td>
<td>4.1</td>
<td>3.88 to 4.31</td>
<td>0.9995</td>
</tr>
</tbody>
</table>

*: EC₅₀ = Concentration of Na₂EDTA (µmol L⁻¹) at which survival was 50% as determined by fitting the experimental data to a four-parameter logistic model (Hill equation) using computer curve-fitting software (Prism 4, GraphPad Software, Inc, San Diego, CA, USA), ²Leaves: 95% Confidence Intervals of fitted EC₅₀. ³Correlation coefficient of fitted curves

Table 3: Production of ochratoxin A by *A. carbonarius* MUM 03.59 and *A. niger* MUM 03.57 in CYA and in CYA supplemented with 1 and 10 mM Na₂EDTA (CYA-1 m and CYA-10 m, respectively)

<table>
<thead>
<tr>
<th>Strain</th>
<th>Days</th>
<th>Medium</th>
<th>CYA</th>
<th>CYA+1 m</th>
<th>CYA-10 m</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>A. carbonarius</em></td>
<td>0</td>
<td>nd</td>
<td>nd</td>
<td>nd</td>
<td>nd</td>
</tr>
<tr>
<td>MUM 03.59</td>
<td>2</td>
<td>1.49±0.29</td>
<td>nd</td>
<td>nd</td>
<td>nd</td>
</tr>
<tr>
<td>4</td>
<td>14.53±3.53</td>
<td>19.42±1.44</td>
<td>nd</td>
<td>39.12±1.54</td>
<td>nd</td>
</tr>
<tr>
<td>6</td>
<td>48.3±4.83</td>
<td>82.3±9.53</td>
<td>nd</td>
<td>0.08±0.11</td>
<td>nd</td>
</tr>
<tr>
<td>8</td>
<td>68.61±5.62</td>
<td>82.3±9.53</td>
<td>nd</td>
<td>0.08±0.11</td>
<td>nd</td>
</tr>
<tr>
<td><em>A. niger</em></td>
<td>0</td>
<td>nd</td>
<td>nd</td>
<td>nd</td>
<td>nd</td>
</tr>
<tr>
<td>MUM 03.57</td>
<td>2</td>
<td>0.12±0.02</td>
<td>nd</td>
<td>nd</td>
<td>nd</td>
</tr>
<tr>
<td>4</td>
<td>1.69±0.28</td>
<td>0.30±0.04</td>
<td>nd</td>
<td>1.68±0.09</td>
<td>nd</td>
</tr>
<tr>
<td>6</td>
<td>10.01±0.58</td>
<td>1.68±0.09</td>
<td>nd</td>
<td>0.20±0.02</td>
<td>nd</td>
</tr>
<tr>
<td>8</td>
<td>11.81±0.59</td>
<td>7.56±0.39</td>
<td>nd</td>
<td>0.20±0.02</td>
<td>nd</td>
</tr>
</tbody>
</table>

Values are the mean of three replicates ± Standard Deviation (SD). For each strain and column, data marked with different letter(s) are significantly different at p < 0.05 for the Duncan's post hoc test, nd: not detected

in 50% (EC₅₀) is 2.1 µmol L⁻¹ for *A. carbonarius*, 0.9 µmol L⁻¹ for *A. ibericus*, 2.0 µmol L⁻¹ for *A. niger* MUM 03.57 and 4.1 µmol L⁻¹ for *A. niger* MUM 03.58 (Table 2).

The production of ochratoxin A was also affected by the presence of Na₂EDTA in culture media. When 1 mM Na₂EDTA was used, OTA was detected after 4 days of growth, while when 10 mM Na₂EDTA were used, it was detected after 8 days (Table 3). The accumulation of OTA was significantly lower in most situations. In the presence of 1 mM Na₂EDTA and after 8 days of growth, *A. niger* MUM 03.57 produced 7.5 µg OTA/plate. After the same period of incubation with 10 mM Na₂EDTA this strain produced only 0.2 µg OTA/plate, which is 98.3% less than the respective control. Under the same conditions, the *A. carbonarius* strain produced only 0.08 µg OTA/plate, 99.9% less than the control. The reduction in OTA production was not observed when *A. carbonarius* was grown with 1 mM L⁻¹ of Na₂EDTA (Table 3).

Several studies have previously reported the bactericidal properties of Na₂EDTA (Ohta, 2003; Reidermiller et al., 2006). However, fungi tolerance to Na₂EDTA is not well documented despite its utilization by the food and agrochemical industry, being its antifungal effects mainly reported on clinical strains as Candida albicans (Sen et al., 2000) or Aspergillus fumigatus (Hachem et al., 2006).

In this study, it was demonstrated that Na₂EDTA can significantly inhibit the growth rate of black aspergilli isolated from grapes and significantly delay and reduce the ochratoxin A produced by the ochratoxigenic strains tested. Namely, 2 mM L⁻¹ of Na₂EDTA were sufficient to reduce the growth rate of ochratoxigenic strains in 50% and 10 mM L⁻¹ of Na₂EDTA to delay the production of OTA in 8 days and reduce the levels produced in approximately 99%. The effect of Na₂EDTA on strains growth is probably due to a defective cell wall construction mediated by its zinc binding capacity as presented by Brul et al. (1997) for yeasts. The reductions on OTA amounts produced are probably due to the inhibition of strains growth further than to the inhibition of the mycotoxin synthesis.
In agriculture, Na₂EDTA is commonly used on foliar products to supplement micronutrients to plants or to stabilize the formulation of several agrochemical products. It will be interesting to study, *in vivo*, if the application of products that contain EDTA can contribute to control the presence of black ochratoxigenic aspergilli and so the levels of ochratoxin A in agriculture commodities such as grapes.

SAFETY

Ochratoxin A is a toxic compound that needs to be manipulated with care and with appropriate safety precautions.

ACKNOWLEDGMENT

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REFERENCES


