

Can integrated prophages affect virulence and fitness of the honeybee pathogen *Paenibacillus larvae*?



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INTRODUCTION

Paenibacillus larvae is a spore-forming Gram-positive bacterium, with five distinct genotypes (ERIC I-V), that causes one of the most destructive bacterial honeybee brood diseases – American Foulbrood (AFB)¹. Temperate phages (prophages) play an important role in the evolution of bacterial populations across all ecosystems, often providing new genes and host genome rearrangements^{2,3,4}. They are, thus, able to alter bacterial phenotypic traits, at fitness and virulence level, or give host protection through superinfection³. So far, the impact of prophages on *P. larvae* ecology has not been evaluated. Our main goal was to understand the potential role or impact of prophages on fitness and virulence of *P. larvae*.

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METHODS

Data collection
 GenBank (accessed April 2020)
 - *P. larvae* genomes (n=14)
 - n=11 unique sequences
 - n=3 sequences excluded
 (2: high no. of contigs; 1: repeated)

Detection and curing of prophages in *P. larvae* strains
 Accession no. for bacterial genomes
 PHASTER output: Intact or defective (incomplete and questionable) prophages

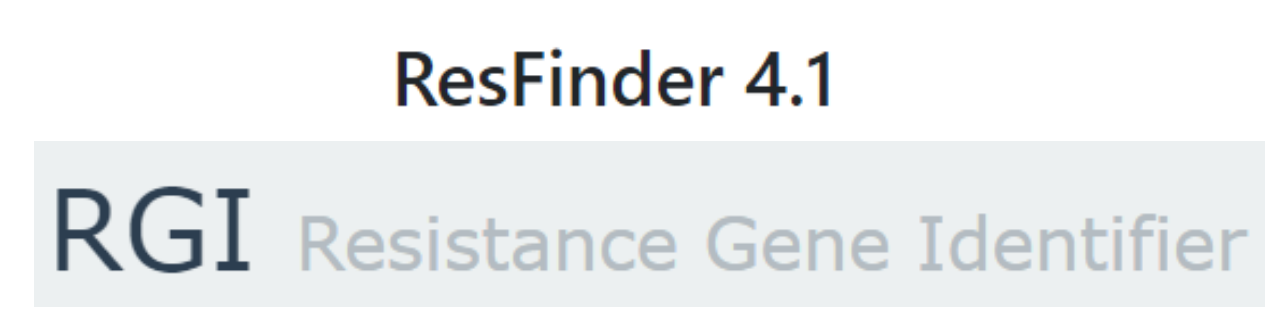
Manual curing:
 - Presence of lysin (endolysin or other)
 - Structural CDSs enough
 - Assembly CDSs (small and large terminase)

BLASTp default and tailed phages (Tax id: 28883)
 CD-Search Tool: E-value cut-off of 1×10^{-5}



Virulence / fitness features provided by prophages
Antibiotic resistance genes:
 - ResFinder
 - Resistance Gene Identifier (RGI) of CARD (The Comprehensive Antibiotic Resistance Database)
 - W/ perfect, strict and loose hits

Adapted COG (Cluster of Orthologous Groups) analysis
 - Proteins grouped according to the function



RESULTS

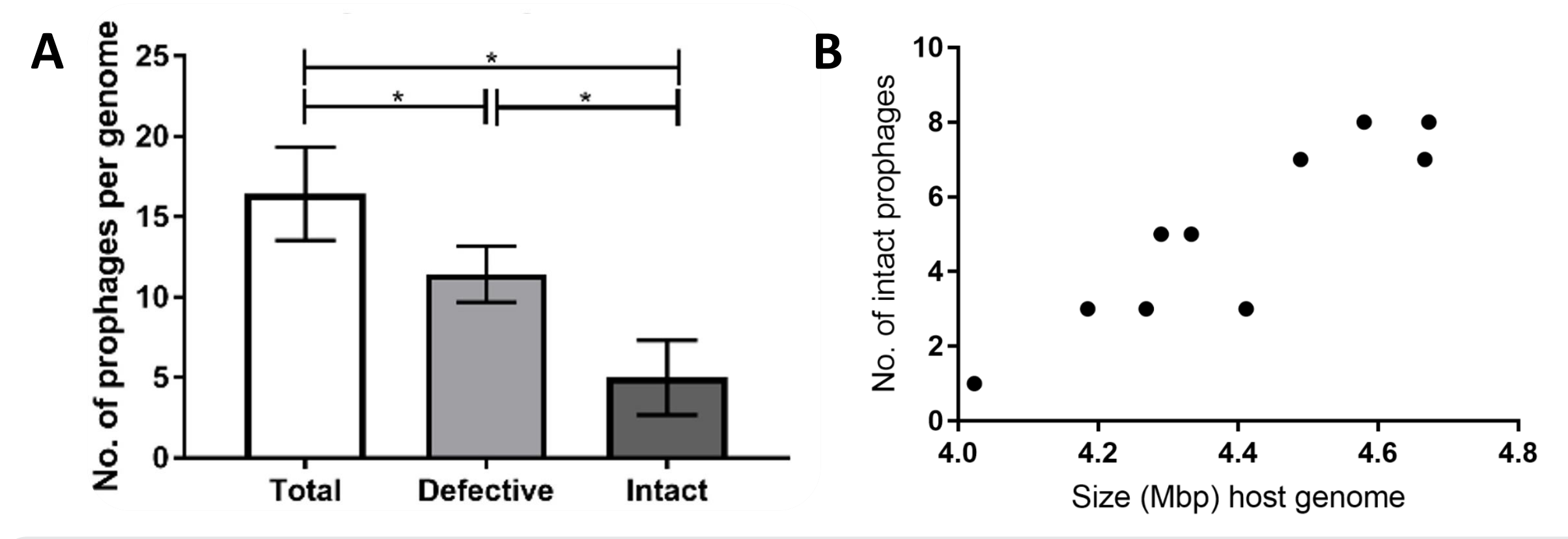


Figure 1. (A) Total, defective and intact prophages/ host genome. (B) No. of intact prophages/ size of host genome.

- Each *P. larvae* genome held 5.0 ± 2.3 intact and 11.5 ± 1.8 defective prophages;
- The prophage frequency of occupation was $5.8 \pm 2.5\%$.
- *P. larvae* with larger genomes harboured a higher number of intact prophages.

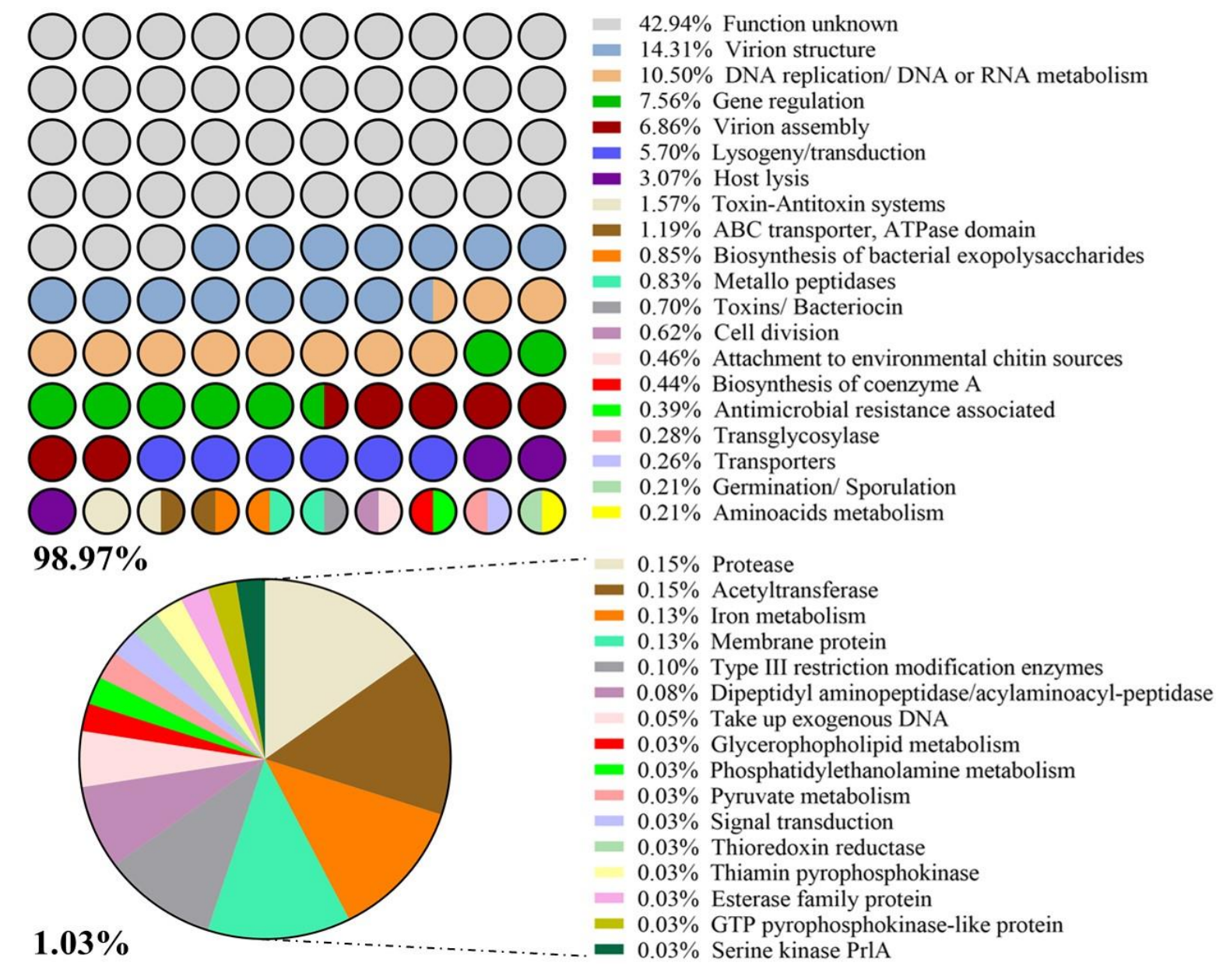


Figure 2. List of COG categories. % of prophage-derived CDS with a given function/ group.

- 3876 prophage proteins.
- 36 functional categories. 95% of proteins ≥ 1 homologous sequence with tailed phages.
- 43% proteins- unknown function.
- Lysogeny/transduction group is 5.7% of the proteins, transposase (n= 112) was the most often identified CDS.
- TA systems: virulence and fitness-related category with the highest % -> 1.6%.

➤ ResFinder / RGI – no functional antimicrobial resistance genes. Yet, TetR family transcriptional regulator, metallo- β -lactamase and β -lactamase inhibitory proteins were identified.

➤ CDS provided by prophages only found in:

Genotype	No. of unique CDS found in intact prophage - e.g. of CDS
ERIC I	21 CDS – Efflux transporter, bacteriocin-like clocisticin, DNA internalization protein ComEC/Rec2, enhancer-like protein, histidine kinase-like protein, pyruvate dehydrogenase E1
ERIC II	5 CDS – Antitoxin SocA, FtsX-like permease, MazG-like nucleotide pyrophosphohydrolase, outer spore coat protein (CotE) and DNA mismatch repair protein MutS
ERIC III	2 CDS – DNA binding YncE and phosphatidylglycerophosphatase
ERIC IV	2 CDS – Iron-sulfur (Fe-S) uptake (SufB) and nitrogen fixation (NifU)
ERIC V	7 CDS – Aromatic acid exporter family, leukocidin subunit LukF-PV precursor, toxins (2), membrane protein, chitin-binding protein GbpA and thioredoxin reductase

CONCLUSIONS

- 30.4% of detected prophages were intact.
- The high number of proteins associated with transport and exchange of genomic DNA fragments can be responsible for prophage and host genomes rearrangements.
- Some CDS were widely distributed across all genotypes, e.g.: HicB and MazE antitoxins, YopX protein.
- For each function associated with virulence and fitness, the percentage of trait was less than 2% in the COG analysis.
- Only ERIC V strains appear to have a competitive advantage since prophages contained multiple CDS that could contribute to a more aggressive infection (LukF-PV and GbpA).

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References
¹Gensch, E. (2010). American Foulbrood in honeybees and its causative agent, *Paenibacillus larvae*. ²Fortier, L. (2017). The Contribution of Bacteriophages to the Biology and Virulence of Pathogenic Clostridia. ³Brussow, H. et al. (2004). Phages and the Evolution of Bacterial Pathogens: from Genomic Rearrangements to Lyso-genic Conversion. ⁴Wachino, J. et al. (2019). Intercellular Transfer of Chromosomal Antimicrobial Resistance Genes between *Acinetobacter baumannii* Strains Mediated by Prophages.