High Enthalpy and Low Enthalpy Death in Saccharomyces cerevisiae Induced by Acetic Acid

Isabel Pinto, Helena Cardoso, and Cecília Leão

Laboratory of Microbiology, University of Minho, 4719 Braga Codex, Portugal

N. van Uden*

Laboratory of Microbiology, Gulbenkian Institute of Science, 2781 Oeiras Codex, Portugal

Accepted for publication June 28, 1988

Acetic acid at concentrations as may occur during vinification and other alcoholic yeast fermentations induced death of glucose-grown cell populations of Saccharomyces cerevisiae IGC 4072 at temperatures at which thermal death was not detectable. The Arrhenius plots of specific death rates with various concentrations of acetic acid (0-2%, w/v) pH 3.3 were linear and could be decomposed into two distinct families of parallel straight lines, indicating that acetic acid induced two types of death: 1) High enthalpy death (HED) predominated at lower acetic acid concentrations (<0.5%, w/v) and higher temperatures; its enthalpy of activation (ΔH^{\star}) approached that of thermal death $(12.4 \times 10^4 \text{ cal/mol})$; 2) Low enthalpy death (LED) predominated at higher acetic acid concentrations and lower temperatures with ΔH^{\neq} of 3.9 \times 10⁴ cal/mol. While the ΔH^{\neq} values for HED induced by acetic acid were similar with those reported earlier for HED induced by other fermentation endproducts, the values for the entropy coefficients were different: 127-168 entropy units mol⁻¹L for acetic acid as compared with 3.6-5.1 entropy units mol⁻¹L for ethanol, which agreed with experimental results indicating that acetic acid is over 30-times more toxic than ethanol with respect to yeast cell viability at high process temperatures.

INTRODUCTION

Fermentation end-products other than ethanol may enhance the apparent toxicity of the latter for *Sacch. cerevisiae* with respect to growth, fermentation and viability.¹⁻³ Acetic acid is a normal end-product of fermentation by this yeast^{3,4} and additional amounts may be produced by contaminating acetic acid bacteria.⁵ Acetic acid is not metabolized by glucose-repressed yeast and enters the cell in the undissociated form by simple diffusion where it dissociates and, if the external pH is lower than the intracellular one, will accumulate as a function of $\Delta p H$.^{6,7}

During fermentation of grape must or other solutions of hexoses (including invert sugars which are rapidly produced when sucrose is the fermentation substrate) the yeast is repressed with respect to acetate metabolism.

Here evidence is presented that under such conditions acetic acid induces two types of death in *Sacch. cerevisiae* at concentrations as may occur during vinification and other alcoholic yeast fermentations.

MATERIALS AND METHODS

Strain

Saccharomyces cerevisiae IGC 4072 was originally isolated from a sample of Fermivin, an industrial wine yeast distributed by Rapidase, Selin, France. It was maintained on slants of glucose (2%, w/v), peptone (1%, w/v), yeast extract (0.5%, w/v), and agar (2%, w/v).

Viability experiments

Loss of viability of *Sacch*. *cerevisiae* populations induced by acetic acid was measured by methods described earlier for alkanol-induced death.⁸

RESULTS AND DISCUSSION

Experimental specific death rates (k_d) obtained at pH 3.3 in the presence and the absence of acetic acid were plotted as modified Arrhenius plots according to the theory of absolute reaction rates^{9,10} (Fig.1):

$$\ln \frac{k_d}{T} = \ln \frac{k_B}{h} + \frac{\Delta S^{\neq}}{R} - \frac{\Delta H^{\neq}}{R} \frac{1}{T}$$
(1)

where T is absolute temperature, k_B is Boltzmann's constant, h is Planck's constant, R is the gas constant, ΔS^{\neq} is the entropy of activation and ΔH^{\neq} is the enthalpy of activation.

The combinations of temperature and concentrations of acetic acid were chosen in such a way that the experimental specific death rates were of the same order of magni-

^{*} Author to whom all correspondence should be addressed.

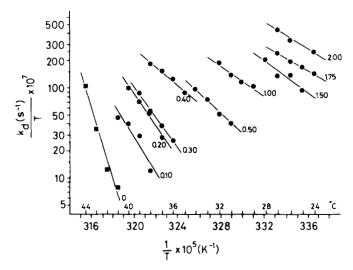
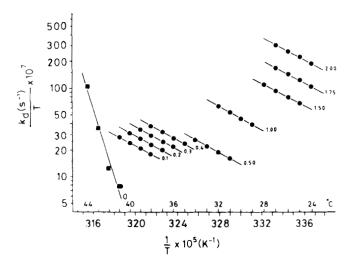


Figure 1. Dependence on the temperature of the specific death rates (k_d) of Saccharomyces cerevisiae IGC 4072 at pH 3.3 in the absence and the presence of acetic acid. Arrhenius plots according to eq. (1). Numbers indicate extracellular concentrations of acetic acid (%, w/v).

tude. Under these experimental conditions the absolute values of the slopes of the plots (i.e., $\Delta H^{\neq}/R$) decreased with increasing concentrations of acetic acid and decreasing experimental temperatures (Fig. 1). This suggested that two different types of death were induced by acetic acid: a high enthalpy death (HED) predominant at higher temperatures and lower acetic acid concentrations and a low enthalpy death (LED) predominant at lower temperatures and higher acetic acid concentrations. A similar behavior was observed earlier in *Sacch. cerevisiae* with respect to ethanol-induced death.¹¹

Theoretical Arrhenius plots of LED (Fig. 2) were calculated using the following form of eq. (1):

$$\ln \frac{k_{d}}{T} = \ln \frac{k_{B}}{h} + \frac{\Delta S_{0}^{\neq} + C_{E}^{A} X}{R} - \frac{\Delta H^{\neq}}{R} \frac{1}{T}$$
(2)



$$\Delta S_X^{\neq} = \Delta S_0^{\neq} + C_E^A X \tag{3}$$

where ΔS_X^{\neq} is the entropy of activation of LED in the presence of concentration X of acetic acid in the medium and C_E^A the respective entropy coefficient (for a discussion of these entropy relations see Leão and van Uden⁸). Values for the different parameters used in the calculations were obtained as follows: ΔH^{\neq} was the average of the experimental ΔH^{\neq} values at pH 3.3 at concentrations 1.5, 1.75, and 2.0% (w/v) acetic acid. An estimate of entropy coefficient C_E^A was obtained from the linear isothermic death plot at 26°C, pH 3.3 (Fig. 4) according to the following equation:⁸

$$\ln k_d^{\chi} = \ln k_d^0 + \frac{C_E^A}{R} X \tag{4}$$

where k_d^0 and k_d^x are the specific death rates in the absence and the presence of concentration X of acetic acid. Using this estimate and an experimental value of ΔS_x^{\neq} obtained at 2% (w/v) acetic acid, an estimate of ΔS_0^{\neq} was calculated by the use of eq. (3).

The theoretical Arrhenius plots of LED thus obtained (Fig. 2) were subtracted from the experimental plots. In this way a family of high enthalpy plots was obtained that represented HED death alone (Fig. 3). The plots were processed as described earlier for alkanol enhanced thermal death⁸ and the following estimates where obtained: $\Delta H^{\neq} = 12.4 \times 10^4$ cal/mol a value similar with ΔH^{\neq} of thermal death (i.e., the value at 0% acetic acid) and C_{ε}^{A} 168 entropy units mol⁻¹ L. A similar estimate for C_{ε}^{A} (127 entropy units mol⁻¹ L) was obtained from an isothermal death plot at 39°C, pH 3.3 (Fig. 4).

It is concluded that HED death induced by acetic acid represented thermal death enhanced exponentially by the acid and that the toxic effect was reflected in the values of

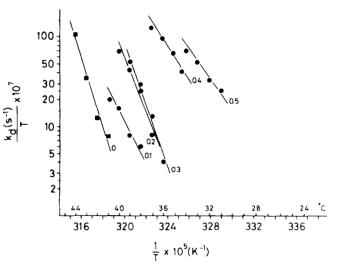


Figure 2. Theoretical Arrhenius plots calculated by the use of eqs. (2) and (3) of low enthalpy death of *Saccharomyces cerevisiae* IGC 4072 induced by acetic acid under the experimental conditions of Figure 1. Numbers indicate extracellular concentrations of acetic acid (%, w/v). (**II**) Experimental thermal death rates.

Figure 3. Theoretical Arrhenius plots, obtained by subtracting the plots of Figure 2 from the plots of Figure 1, of high enthalpy death in *Saccharomyces cerevisiae* IGC 4072 induced by acetic acid. Numbers indicate extracellular concentrations of acetic acid (%, w/v). (\blacksquare) Experimental thermal death rates.

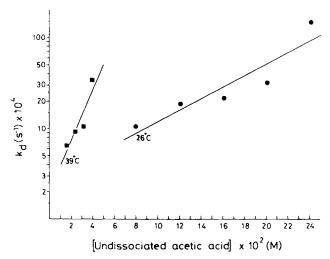


Figure 4. Dependence on the concentrations of extracellular acetic acid of the specific death rates (k_d) of Saccharomyces cerevisiae IGC 4072 at constant temperature and pH 3.3.

 ΔS^{\neq} but not of ΔH^{\neq} (parallel Arrhenius plots). Furthermore acetic acid induced LED with similar characteristics but distinct from thermal death by its much lower ΔH^{\neq} (3.9 × 10⁴ cal/mol) and C_{E}^{A} (24 entropy units mol⁻¹ L).

HED is of practical importance since it contributes to so-called "heat-sticking" of alcoholic yeast fermentations particularly of red wine and fuel ethanol fermentations in warm countries in the absence of efficient temperature control.¹² As reported earlier ethanol and other alkanols⁸ as well as octanoic and decanoic acids² also induced this type of cell death. While in all these cases the ΔH^{\neq} values were similar with ΔH^{\neq} of thermal death, the entropy coefficients had vastly different values (Table I). Introduction in eq. (4) of the entropy coefficients for HED induced by ethanol and by acetic acid led to the prediction that the latter is over 30-times more toxic, with respect to yeast cell viability at high process temperatures, than the latter (on a molar basis). This prediction is in agreement with experimental results (compare Figure 3 of the present article with Figure 1 of Leão and van Uden⁸). Thus these two fermentation endproducts may both contribute to significant HED at realistic concentrations and high process temperatures. A similar reasoning applies to the much more toxic octanoic and decanoic acids.²

With respect to LED acetic acid is also more toxic than ethanol (compare Figure 1 of the present article with Fig-

Table I. Entropy coefficients of the enhancement of high enthalpy death (thermal death) in *Saccharomyces* by alkanols and fatty acids.

| Alkanols Fatty acids | Entropy coefficient (entropy units $mol^{-1}L$) | Species | Reference |
|-------------------------|--|-------------------|-----------------|
| Ethanol | 3.6–5.1 | Sacch. cerevisiae | 8 |
| Isopropanol | 9.6-10.2 | Sacch. cerevisiae | 8 |
| Propanol | 15.3-17.4 | Sacch. cerevisiae | 8 |
| Butanol | 58.3-60.0 | Sacch. cerevisiae | 8 |
| Octanoic acid | 10.3×10^{3} | Sacch. bayanus | 2 |
| Decanoic acid | 31.6×10^{3} | Sacch. bayanus | 2 |
| Acetic acid | 127-168 | Sacch. cerevisiae | present article |

ure 2 of Sá-Correia and van Uden¹¹). However, significant LED at intermediate and low temperatures requires rather high concentrations of the toxic metabolites which at least in the case of acetic acid are less realistic than the concentrations of acetic acid that induce significant HED at high process temperatures.

It remains to be seen which form(s) of acetic acid (acetate ion, undissociated acid or both) is(are) involved in HED and LED.

I. P., H. C., and C. L. were supported by a research grant from INIC, Lisbon, Portugal.

References

- S. Lafon-Lafourcade, C. Geneix, and P. Ribéreau-Gayon, Appl. Environ. Microbiol., 47, 1246 (1984).
- 2. I. Sá-Correia, Biotechnol. Bioeng., 28, 761 (1986).
- M. N. Pons, A. Rajab, and J. M. Engasser, Appl. Microbiol. Biotechnol., 24, 193 (1986).
- 4. R. B. Beelman and J. F. Gallander, Adv. Food Res., 25, 1 (1979).
- 5. A. Joyeux, S. Lafon-Lafourcade, and P. Ribéreau-Gayon, Sciences Aliments, 4, 247 (1984).
- 6. C. Leão and N. van Uden, Appl. Microbiol. Biotechnol., 23, 389-393 (1986).
- F. Cássio, C. Leão, and N. van Uden, *Appl. Environ. Microbiol.*, 53, 509 (1987).
- 8. C. Leão and N. van Uden, Biotechnol. Bioeng., 24, 1581 (1982).
- F. H. Johnson, H. Eyring, and M. J. Polissar, *The Kinetic Basic of Molecular Biology* (John Wiley and Sons, New York, 1954).
- 10. N. van Uden, Adv. Microb. Physiol., 25, 195 (1984).
- 11. I. Sá-Correia and N. van Uden, *Biotechnol. Bioeng.*, 28, 301-303 (1986).
- 12. N. van Uden, Ann. Rep. Ferment. Proc., 8, 11-58 (1985).