Cork extracts exhibit thermo-oxidative protection properties in polypropylene–cork composites and as direct additives for polypropylene

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ABSTRACT

The thermo-oxidative stability of polypropylene (PP) in composites containing 15 wt.% of cork and the performance of selected cork extracts as stabilizing additives for PP was evaluated by Oxidation Induction Time (OIT) and by Oxidation Onset Temperature (OOT). The results showed that cork increases the OIT of PP in the composite and it was identified that the cork extractives fraction is responsible for such behavior. Selected cork extracts with high antioxidant capacity (determined by dpph radical scavenging and oxygen reactive absorbance capacity assays) were compounded by extrusion with PP in 0.5 and 1.5 wt.%. It was found that the ethanol extract is the most effective as thermo-oxidative stabilizer for PP. At the loading level of 1.5%, the OIT increases from 3.8 (neat PP) to 29.7 min at 200 °C and from 1.2 (neat PP) to 9.0 min at 220 °C. The OOT also increases from 216 °C (neat PP) to 247 °C. Mechanical tests, performed on PP loaded with the cork extracts, showed that the presence of these extracts has no significant effect on the polymer mechanical performance. The results demonstrate the suitability of cork as a source of thermo-oxidative stabilizing additives for the formulation of polyolefins, and enable the exploitation of new routes of cork valorization.

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1. Introduction

Cork is the outer bark of the oak tree Quercus suber L., which mainly grows in western Mediterranean regions. The bark is manually harvested every 9–12 years in a process that does not damage the tree, allowing it to continue to develop normally. Europe holds ca 60% of the total area of cork oak forests and harvests more than 80% of the World’s cork. Portugal is the major producer, being responsible for the processing of 49% of the total amount of cork harvested in the World [1,2].

The cork chemical composition is variable and is dependent on different factors, such as, geographical origin, climate, age of the trees, etc. Anyway, typical compositions are within the ranges: 25–40% for suberin; followed by 13–29% for lignin; 6–25% for polysaccharides; and 8–24% for extractives [3–5]. Structurally, cork is constituted by an arrangement of cells (that have lost its internal content) with highly suberized walls. When observed on the microscope, this structure resembles a honeycomb pattern. This material presents some remarkable properties, such as: near-zero Poisson coefficient; complete recovery of shape after compression; among many others [6–8]. These properties are devised from its unique chemical composition and morphological structure.

The main industrial application of cork is for the production of stoppers. However, there are large quantities of cork (in different forms), whose lower quality or format limitation, does not allow its processing for the preparation of stoppers. This large amount of unused cork (usually considered as a by-product) has prompted the development of other applications for this unique natural material. In this scope, a series of applications for cork and cork-based materials have been developed; these include the production of: lightweight, sound and vibration insulation materials for housing and aerospace applications; cork–rubber composites for...
automobile engine gaskets; or decorative items. Despite these alternative uses, up to 25% of the processed cork is still transformed into powder that has no direct application. Therefore, the primary cork constituents are responsible for the above-mentioned thermo-oxidative protection. Moreover, we have evaluated if these can be further valorized as direct stabilizing agents for PP and we have quantified the protective effect by oxidation induction time (OIT) and oxygen onset temperature (OOT) parameters.

2. Materials and methods

2.1. Materials and reagents

Cork powder was selected and collected at the industrial facilities directly from the processing machines and used as received (Amorim Cork Composites, Portugal). This material is produced by reprocessing of cork scrapes from different origins. The leftovers are collected, further milled and sorted by density and particle size in a high frequency industrial vibrating equipment, to obtain homogenous fractions, before further industrial use. From the different fractions arising from this process, for this work, the criterion for selecting the cork powder sample was the availability in large quantities and the purity (absence of other contaminants). The selected cork powder was collected at the industrial facility during the normal production process, and was considered to be representative of the daily operations by the industry responsible.

The matrix was a PP homopolymer (1374 E2, ExxonMobil, Germany), with an MFI of 20.8 g 10 min$^{-1}$ (200 °C, 2.16 kg), and a melting point of 162 °C, that was transformed into powder and supplied by Pallmann Maschinenfabrik GmbH & Co. (Germany). All chemical reagents were analytical grade. Ethanol and chloroform were purchased from Fisher Chemicals, while dichloromethane, methanol sulfuric acid, Pentaerythritol tetrakis[3-(3,5-di-tert-butyl-4-hydroxyphenyl)propionate], Folin–Ciocalteu reagent, gallic acid, 2,2-diphenyl-1-picrylhydrazyl, 2,2-azobis(2-methylpropionamide)dihydrochloride, gallic acid, 2,2-diphenyl-1-picrylhydrazyl, 2,2-azobis(2-methylpropionamide)dihydrochloride, fluorescein sodium salt and 6-hydroxy-2,5,7,8-tetramethylchromane-2-carboxylic acid (Trolox) were purchased from Sigma–Aldrich. Sodium hydroxide and sodium carbonate were acquired from Fluka.

2.2. Cork powder characterization and preparation of extractive-free cork powder

The cork powder particle size distribution was determined on an analytical sieve shaker (model AS200, Retsch, Germany) equipped with a cascade of different mesh sieves. The chemical composition was determined in terms of extractives, suberin and lignin, following already established protocols reported elsewhere [3,35]. The extractive fraction was quantified as the combined mass of the material solubilized and recovered after successive Soxhlet extractions with dichloromethane, ethanol and water, for periods of 6 h each. The resulting cork powder was defined as extractives-free cork powder and was further used for the preparation of composites (hereafter designated by PP–Cork–EF) and for the suberin content determination. The suberin content was quantified as the chloroform soluble fraction after submitting the extractive-free cork powder to alkaline methanolysis. In short, the cork material was kept under reflux in methanol containing 0.1 M sodium hydroxide (1 g per 50 mL), for 6 h, after which, the liquid fraction was recovered, acidified to pH 6 with hydrochloric acid; after solvent removal, the solid residue was suspended in water and extracted with chloroform. The suberin material was obtained by removal of the chloroform under vacuum in a rotatory evaporator. The lignin fraction was obtained from the dessuberized cork material by processing it with 72% sulfuric acid in a 15 mL per 1 g of cork ratio, at room temperature, under constant stirring. After 2 h, 30 mL of concentrated sulfuric acid per gram of material was added and the mixture refluxed for 4 h. After thorough washing with distilled
water, the lignin material was recovered as the enduring solid residue.

2.3. Cork extracts and antioxidant properties

Cork extracts were obtained by direct hot solvent extraction from cork powder. The extracts were obtained by maintaining cork powder in contact with the selected solvents (1 g per 50 mL) under reflux conditions, for a period of 6 h. After cooling, the liquid fraction was recovered by filtration and concentrated by vacuum evaporation. After freeze-drying, the extracts were recovered in the form of powders, stored in a desiccator and protected from light until further use. The extracts were obtained with water, ethanol and water:ethanol 1:1 (v/v) and are hereafter designated E.H2O, E.EtOH and E.EtOH50, respectively.

The cork extracts were characterized for its total phenolic content (TPC) and its antioxidant potential as scavengers of radicals and reactive oxygen species. The TPC was determined by the Folin–Ciocalteu assay; the radical scavenging capacity by the 2,2'-diphenyl-1-picrylhydrazyl radical scavenging assay (dpph assay) and the reactive oxygen species scavenging by the oxygen radical absorbance capacity (ORAC) assay.

The TPC was determined according to the Folin–Ciocalteu method [36] and adapted from Santos et al. [37]. Briefly, 40 µL of a suitable sample dilution were added to 500 µL of diluted Folin reagent (1:10 with water) and 460 µL of a 75 g/L aqueous sodium carbonate solution. The reaction mixtures were kept at 50 °C for 5 min and, after cooling, 250 µL were transferred to individual wells of a 96 well microplate and the absorbance was read at 760 nm (microplate reader Synergy HT, Bio Tek, USA). The TPC was calculated from a standard curve with gallic acid (range 1.5–20 µg/mL) and expressed as mg of gallic acid equivalents per gram of extract, mgGAeq/g Extract.

The dpph radical scavenging activity was determined following a methodology adapted from by Santos et al. [37]. Briefly, 35 µL of suitable dilutions of extract solutions were added to 900 µL of a solution of dpph in methanol with initial absorbance of 0.850 ± 0.01. After vigorous vortexing the reaction solutions were kept in the dark for 30 min. The absorbance of 200 µL aliquots was read at 517 nm (microplate reader Synergy HT, Bio Tek, USA). The EC50 was calculated from the plot of scavenging activity against concentration and represent the amount of extract necessary to decrease the initial dpph concentration by 50%.

The ORAC antioxidant test procedure was adapted from the methodology described by Huang et al. [38]. The reaction mixture was obtained by mixing: 25 µL of sample solution with 25 µL of 250 mM AAPH (2,2'-Azobis(2-methylpropionamide)dihydrochloride) solution in PBS and 150 µL of a 0.025 µM fluorescein solution. Fluorescein measurements were performed in a microplate reader (Synergy HT, Bio Tek, USA) equipped with adequate fluorescence filters. The white opaque 96 well plaque was maintained at 37 °C, with readings taken at 2 min interval with agitation in between. Fluorescence conditions were excitation at 485 nm and emission at 520 nm. The ORAC value was calculated as the mass of Trolox equivalents per mass of extract, TE.

2.4. Twin-screw extrusion compounding

Prior to compounding, all natural raw materials were dried under vacuum at 30 °C during 24 h to stabilize the cork moisture content. The compositions: neat PP, PP–cork powder composites and PP–Cork extracts, were compounded in a Rondol SCF modular co-rotating twin-screw extruder with the screws diameter of 16 mm, a length to diameter ratio (L/D) = 25 and a single strand die of 3 mm. All processed compositions are discriminated in Table 1, including the temperature profile along the barrel to the die and the rotation screws. The PP–Cork extracts were compounded in a two-step procedure. The extracts were initially solubilized/suspended in ethanol and added to the PP powder in order to obtain the desired loading levels. After thorough mixing, the mixture was dried under vacuum at 30 °C for 24 h to guarantee complete solvent removal. The resulting mixture was then extruded according the conditions indicated in Table 1. The extruded was cooled in a water bath, pelletized in an SF SCHEER & CIE (Germany) equipment and dried at 30 °C under vacuum during 24 h. The final materials were stored in polyethylene bags until further use.

2.5. Thermal properties

The thermal-oxidative properties were accessed by means of OIT and OOT tests determined using a differential scanning calorimeter (DSC, T. A. Instruments, model DSCQ100, USA), with sample mass of 7–9 mg in open aluminum pans. For the OIT determination, samples were heated to the test temperature (200 and 220 °C) under nitrogen atmosphere and stabilized for 2 min. Purge gas was commutted to oxygen and the heat flow vs time was recorded. The test was considered completed on the observation of the exothermal peak caused by the samples oxidation. The OIT value was determined from the time elapsed between the swich to oxygen purge gas and the onset point between baseline and oxidation curve. The OOT determination was performed by equilibrating the temperature to 20 °C before changing the purge gas to oxygen, followed by temperature increase at the rate of 10 °C min⁻¹, recording the heat flow vs temperature. The assay was considered complete upon the observation of the exothermic oxidation peak. The OIT value was determined from the temperature of the onset of the oxidation process. DSC analysis of cork extracts was performed using aluminum pans with lids and 7–10 mg of sample mass, from 20 °C to 200 °C at 10 °C/min, under oxygen atmosphere.

2.6. Mechanical properties

The PP–cork extracts pellets were processed in boards by compression molding, using the conditions from a previous study [12]. Tensile specimens with a dog bone shape, gauge length of 20 mm and cross section (4 × 2 mm) were cut from molded boards using a computer numerical control CNC machine (Roland 3D Plotter MDX-20, UK). The tensile properties of the PP–Cork extracts were measured using an Instron 4505 Universal Machine (USA). The tests were conducted using a 1 kN load cell and a crosshead speed of 5 mm min⁻¹ until rupture. The average and standard deviations were determined using 5 specimens per condition.

3. Results and discussion

3.1. Cork powder characterization

The cork powder sample was characterized in terms of the particle size distribution and for its chemical composition. The particle size distribution analysis and the chemical composition results are presented in Table 2.

The results show that the cork powder is constituted by a narrow range of particle sizes, concentrated within two of the ranges analyzed, being the higher amount (84%) between 0.25 and 0.5 mm and a smaller percentage (15%) between 0.125 and 0.25 mm. This particle size distribution reflects the screening process employed in the industrial facility.

The determination of the cork’s chemical composition revealed that it is constituted by approximately 12% extractives, 33% suberin and 25% lignin, with the remaining being mainly polysaccharides.
and inorganic fractions. The determined values are in accordance with the ones reported by other authors and summarized by Silva et al. [4]; they are also within the range of the intrinsic variability that is present for this natural material.

3.2. PP–cork composites

In accordance with Table 1, two PP–cork composite formulations were prepared (one with untreated cork powder and another one with extractives-free cork powder). The thermo-oxidative properties of these formulations were assessed by the OIT at the temperature of 200 °C. The DSC curves and the values determined for the OIT are shown in Fig. 1.

From the DSC OIT curves, it is apparent that the neat PP, in the absence of stabilizers, is prone to rapid oxidation when submitted to high temperature and an oxidative atmosphere. Upon the switch of the purge gas to molecular oxygen, the heat flow curve directs to negative values (resulting from the exothermal oxidation process) at increasing rate with the establishing of an accentuated exothermal peak in very short time.

The OIT curve for the PP–cork composite formulations, after the switch of the purge gas to oxygen, shows a flat area that extends up to almost 40 min within the total time of the assay, resulting in an OIT value of 30.3 ± 0.3 min. The almost zero value for the variation of the heat flow in the referred region is a result of the absence of thermal processes. When comparing the DSC curves and OIT values for the PP–cork composite and the neat PP we can conclude that the presence of the cork material is imparting an extra stabilization to the composite and, consequently, also to the PP. This result is in agreement with a recent study by Fernandes et al. [12] where it was shown that the cork used in the PP–Cork composites induces an increase of the OIT. Reducing the cork particle size, and consequently increasing the surface area of the cork particles, a higher thermal-oxidative protection of the PP matrix was achieved [12]. It is then reasonable to assume that within the cork material composition there are molecules and/or molecular structures that are able to function as thermo-oxidative stabilizers.

The proposed mechanism for PP oxidation involves an initiation step of formation of primary alkyl radicals through the homolytic cleavage of a carbon–hydrogen bond. This leads to PP degradation which can be further enhanced by the presence of oxygen [30]. Hence, for PP, a stabilizing system will have to act at two levels, scavenging of radicals and quenching of reactive oxygen species. In cork, the extractive’s fraction is composed by low molecular weight molecules, e.g. polyphenols, which are known to present the capacity to scavenge radicals [37,39]. In this context, we hypothesize that these molecules are the responsible for the stabilizing effect observed for PP–cork composites.

The preparation of extractives-free cork powder was achieved through the selective removal of this fraction by a procedure employing 3 sequential Soxhlet extractions with dichloromethane, ethanol and water. The extractives-free cork powder was used to obtain a PP–cork EF composite, using the processing conditions previously described. The OIT curve obtained for such composite is also shown in Fig. 1. When the extractives fraction is removed, the

<table>
<thead>
<tr>
<th>Sample</th>
<th>Components</th>
<th>Processing conditions</th>
</tr>
</thead>
<tbody>
<tr>
<td>PP</td>
<td>100</td>
<td>100; 170; 180; 185</td>
</tr>
<tr>
<td>PP–cork</td>
<td>85</td>
<td>80</td>
</tr>
<tr>
<td>PP–cork–EF</td>
<td>85</td>
<td>15</td>
</tr>
<tr>
<td>E_H2O-0.5%</td>
<td>99.5</td>
<td>0.5</td>
</tr>
<tr>
<td>E_H2O-1.5%</td>
<td>98.5</td>
<td>1.5</td>
</tr>
<tr>
<td>E_EtOH-0.5%</td>
<td>99.5</td>
<td>0.5</td>
</tr>
<tr>
<td>E_EtOH50-0.5%</td>
<td>99.5</td>
<td>0.5</td>
</tr>
<tr>
<td>E_EtOH50-1.5%</td>
<td>98.5</td>
<td>1.5</td>
</tr>
<tr>
<td>IR-0.5%</td>
<td>PP with 0.5% Pentaerythritol tetrakis(3,5-di-tert-butyl-4-hydroxyphenyl)propionate</td>
<td></td>
</tr>
</tbody>
</table>

Table 1
Compositions and processing conditions used on the preparation of the PP–cork composites and PP–cork extracts.

<table>
<thead>
<tr>
<th>Sample</th>
<th>Components</th>
<th>Processing conditions</th>
</tr>
</thead>
<tbody>
<tr>
<td>PP</td>
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<td>PP–cork</td>
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<td>85</td>
<td>15</td>
</tr>
<tr>
<td>E_H2O-0.5%</td>
<td>99.5</td>
<td>0.5</td>
</tr>
<tr>
<td>E_H2O-1.5%</td>
<td>98.5</td>
<td>1.5</td>
</tr>
<tr>
<td>E_EtOH-0.5%</td>
<td>99.5</td>
<td>0.5</td>
</tr>
<tr>
<td>E_EtOH50-0.5%</td>
<td>99.5</td>
<td>0.5</td>
</tr>
<tr>
<td>E_EtOH50-1.5%</td>
<td>98.5</td>
<td>1.5</td>
</tr>
<tr>
<td>IR-0.5%</td>
<td>PP with 0.5% Pentaerythritol tetrakis(3,5-di-tert-butyl-4-hydroxyphenyl)propionate</td>
<td></td>
</tr>
</tbody>
</table>

Table 2
Cork powder particle size distribution and chemical composition.

<table>
<thead>
<tr>
<th>Particle size distribution (% m/m)</th>
<th>Chemical composition (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>&gt;1 mm</td>
<td>12.4 ± 0.7</td>
</tr>
<tr>
<td>&gt;0.5 mm</td>
<td>3.1 ± 0.7</td>
</tr>
<tr>
<td>&gt;0.25 mm</td>
<td>2.9 ± 0.5</td>
</tr>
<tr>
<td>&gt;0.125 mm</td>
<td>6.1 ± 0.3</td>
</tr>
<tr>
<td>&gt;0.063 mm</td>
<td>33.6 ± 1.2</td>
</tr>
<tr>
<td>Tray</td>
<td>25.5 ± 1.8</td>
</tr>
<tr>
<td>Others (polysaccharides, inorganics, etc.)</td>
<td>= 28 (from residue)</td>
</tr>
</tbody>
</table>

DCM: dichloromethane; EtOH: ethanol.

Fig. 1. Determination of the OIT (200 °C) for the PP–Cork composites and the neat PP.
stabilizing effect provided by the cork powder is significantly reduced, reaching an OIT value of only 7.7 ± 0.2 min (which compares to 30.3 ± 0.4 min for the untreated PP–cork). The reminiscent protective effect can be the result of incomplete removal of the extractive fraction and/or antioxidant activity provided by other cork constituents. In fact, Conde et al. [40] have shown that lignin–derived fractions can have some protective effect. Nevertheless, our data clearly indicate that the protective effect arises primarily from the extractive fraction.

3.3. Cork extracts and antioxidant properties

The cork extractive’s fraction is composed of low molecular weight molecules that aren’t covalently bonded to the structural elements of the cell wall (suberin and lignin). Therefore, they are easily extracted using suitable solvents. Partitions of the cork extractives were obtained through direct extractions using water, ethanol and its mixture in a 1:1 ratio.

Such extracts were found to possess high phenolic content and are, therefore, premium candidates for the observed thermo-oxidative stabilization of the polymer in the cork–PP composites. The results for the characterization of the TPC, and the radicals (dpph assay) and reactive oxygen species (ORAC assay) scavenging properties of such extracts are summarized in Table 3, alongside with the yields of extraction.

The results show that the TPC is higher for E_EtOH50 and lower for E_EtOH. When comparing the TPC results with the values from the dpph assay, recalling that these are expressed as an efficient concentration (and, thus, the lower this concentration is, the more effective it is the extract), it can be observed that the radical scavenging capacity is proportional to the TPC of the extract. Indeed, E_EtOH50 is the most effective (5.32 μg/mL) extract for radicals scavenging and E_EtOH (9.44 μg/mL) is the least effective. The ORAC assay measures the extract effectiveness in the absorption of oxygen radical species and, as for the dpph evaluation, the results show that E_EtOH50 (2.11 TE) is the extract with highest activity followed by E_EtOH (1.88 TE). The results for the ORAC assay are consistent with the higher anti-oxidant performance of E_EtOH50, although, in this assay the E_EtOH had a better performance than the E_H2O. This latter variation confirms that the performance of the different extracts is dependent on the nature of the constituent molecules. The water and ethanol cork extracts are composed of numerous molecules with complex structures and is not completely known. However, different phenolic acids, ellagitanins and flavonoids have been identified [37,39].

3.4. Polypropylene—cork extracts

The cork extracts E_H2O, E_EtOH and E_EtOH50 were processed as additives for the stabilization of PP at two loading levels, i.e. 0.5 and 1.5 wt.%. The mixtures were prepared by solubilizing/suspending the selected extracts in ethanol to facilitate its dispersion within the PP powder. The mixtures were processed by extrusion in order to obtain homogenous distribution of the components. The processing conditions employed were the same as described for the processing of PP–cork composites.

The thermal properties of the cork extracts, before processing, were assessed by DSC analysis under oxygen atmosphere. The results are presented in Fig. 2. For lower temperatures, the thermograms present an endothermic band, from approximately 30 °C to 120 °C. The most relevant feature is the onset of an oxidation process (exothermal process) observed for higher temperatures, namely above the 180 °C. During processing, temperatures of 180 °C (and 185 °C for the dye) are used, but the extracts residence time at this temperature will be short and the O2 present will be limited. Therefore, it is not expected the occurrence of significant degradation of the extract during the processing stage.

After being transformed into pellets (of approx. 5 mm in length) and dried, the processed mixtures were tested for its thermo-oxidative stabilization performance. Images depicting the visual aspect of the processed PP loaded with the cork extracts can be found in Fig. 3, for a loading level of 1.5%.

The performance of cork extracts as thermo-oxidative stabilizing agents was assessed by measurement of the OOT and the OIT. The obtained results are presented in Table 4 and the DSC curves for the OOT and the OIT assays are presented in Figs. 4 and 5, respectively. For comparison purposes, a sample of PP containing 0.5% loading of a commercially available hindered phenolic antioxidant (i.e. Pentaerythritol tetrakis(3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate) also known commercially as Irganox 1010) was also compounded and analyzed.

The results clearly demonstrate that the E_EtOH presents the highest performance as a stabilizing additive for PP; the E_H2O is the least effective followed by the E_EtOH50.

The E_H2O, in both tests, OOT and OIT, has a performance that compares to the raw PP. That is, the presence of the extract does not contribute to the protection of the polymer, even considering the 1.5% loading level. The E_EtOH50 shows improved results when compared to the E_H2O, with higher OOT and OIT values. However, this improvement is only marginal when compared to the raw PP. Nevertheless, the E_H2O, in both tests, OOT and OIT, does not have a performance that compares to the raw PP. That is, the presence of the extract does not contribute to the protection of the polymer, even considering the 1.5% loading level. The E_EtOH50 shows improved results when compared to the E_H2O, with higher OOT and OIT values. However, this improvement is only marginal when compared to the raw PP and to other extracts. The OOT is 222.1 °C for 0.5% and 223.9 °C for

<table>
<thead>
<tr>
<th>Extract</th>
<th>Yield, %/m</th>
<th>TPC, mgGAE/gExtract</th>
<th>dpph, EC50 μg/mL</th>
<th>ORAC, TE</th>
</tr>
</thead>
<tbody>
<tr>
<td>E_EtOH</td>
<td>5.8 ± 0.4</td>
<td>396 ± 17</td>
<td>9.44 ± 0.54</td>
<td>1.88 ± 0.20</td>
</tr>
<tr>
<td>E_EtOH50</td>
<td>9.3 ± 0.2</td>
<td>570 ± 39</td>
<td>5.32 ± 0.45</td>
<td>2.11 ± 0.24</td>
</tr>
<tr>
<td>E_H2O</td>
<td>6.7 ± 1.0</td>
<td>486 ± 6</td>
<td>6.33 ± 0.38</td>
<td>1.21 ± 0.25</td>
</tr>
</tbody>
</table>

Fig. 2. DSC analysis of cork extracts under oxygen atmosphere (10 °C/min).

The table above shows the yield, total phenolic content (TPC), dpph radical scavenging and oxygen radical absorbance capacity of cork extracts.
1.5% which compares to 216.2 °C for the neat PP. The OIT is 6.83 min and 7.01 min for 0.5% and 1.5%, which represent a small increase over the value determined for the neat PP (3.83 min). Also of notice (from the results for the E_H2O and E_EtOH50) is that, even with an increase of the loading percentage, the OOT and OIT suffer only a marginal improvement.

In contrast, it is observed a significant improvement of the OOT values for the PP loaded with the E_EtOH, when compared with the neat PP. The temperature of oxidation, for 0.5% and 1.5% loading levels was 241.7 °C and 246.9 °C, respectively. These values compare with 216.2 °C for the neat PP and 247.5 °C for the PP loaded with the commercial antioxidant. The OIT at 200 °C was determined to be 24.28 min for 0.5% and 29.72 min for 1.5%, significantly higher than 3.83 min observed for the neat PP. Anyway, it represents, approximately, 40% and 50% of the commercial antioxidant performance (sample IR-0.5% in Table 4). The OIT values for 220 °C show, as expected, a strong decrease from the ones registered for 200 °C. The OIT of the PP loaded with E_EtOH, at 220 °C was measured to be 6.55 min and 9.00 min for the 0.5% and 1.5% loading, respectively. Indeed, these values represent 55% and 75%, of the performance determined for the commercial antioxidant. The increase in the testing temperature to 220 °C leads to an improvement of the relative performance of the E_EtOH over the commercial antioxidant.

Overall, the results obtained for OOT and OIT show that, for the selected cork extracts, the E_EtOH was the only one who provided effective protection against the thermo-oxidation of PP. This observation is in contrast with what was expected for the

<table>
<thead>
<tr>
<th>Mixture</th>
<th>OOT (°C)</th>
<th>OIT @200 °C (min)</th>
<th>OIT @220 °C (min)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Neat PP</td>
<td>216.2 ± 0.8</td>
<td>3.83 ± 0.08</td>
<td>1.18 ± 0.40</td>
</tr>
<tr>
<td>E_H2O-0.5%</td>
<td>215.7 ± 0.1</td>
<td>3.83 ± 0.13</td>
<td>–</td>
</tr>
<tr>
<td>E_H2O-1.5%</td>
<td>216.0 ± 0.5</td>
<td>3.80 ± 0.10</td>
<td>–</td>
</tr>
<tr>
<td>E_EtOH-0.5%</td>
<td>241.7 ± 0.7</td>
<td>24.28 ± 0.17</td>
<td>6.55 ± 0.11</td>
</tr>
<tr>
<td>E_EtOH-1.5%</td>
<td>246.9 ± 0.7</td>
<td>29.72 ± 0.51</td>
<td>9.00 ± 0.01</td>
</tr>
<tr>
<td>E_EtOH50-0.5%</td>
<td>222.1 ± 1.1</td>
<td>6.83 ± 0.13</td>
<td>–</td>
</tr>
<tr>
<td>E_EtOH50-1.5%</td>
<td>223.9 ± 0.2</td>
<td>7.01 ± 0.52</td>
<td>1.77 ± 0.10</td>
</tr>
<tr>
<td>IR-0.5%</td>
<td>247.5 ± 0.5</td>
<td>60.31 ± 3.10</td>
<td>11.91 ± 0.31</td>
</tr>
</tbody>
</table>

Fig. 3. Images of pellets of neat PP and PP loaded with cork extracts (at 1.5%).

Fig. 4. OOT (°C) determined for neat PP and for the polymer loaded with the E_EtOH50, E_H2O (1.5%) and E_EtOH (0.5% and 1.5%).

Fig. 5. OIT (200 °C) DSC curves for neat PP and for PP loaded with the E_EtOH50, E_H2O (1.5%) and E_EtOH (0.5% and 1.5%).
antioxidant properties of the tested extracts, where the E_EtOH50 presents the highest performance both at the radical scavenging and quenching of oxygen reactive species.

A possible explanation for this observation may reside on the type of molecules that are extracted by each solvent. The extracts obtained with water and ethanol:water (50:50) are constituted by molecules of higher polarity than those present on the ethanol extract, reflecting the properties of the extracting medium. PP is non-polar which can result in incompatibility between the polymeric matrix and these extracts. During the oxidative process, the radicals formed within the polymeric material must be rapidly transferred to the antioxidant systems for stabilization and to avoid the propagation of the chain breaking reactions. Therefore, the antioxidant molecules must be deeply embedded within the polymer matrix for an effective function. If a barrier subsists, the antioxidant system performance will be severely compromised. Therefore, despite of the lower antioxidant capacity revealed, the ethanol extract molecules are more effective in the oxidative protection of PP. Further studies will be required to fully elucidate this possibility.

3.5. Mechanical properties

The mechanical performance of PP loaded with the different cork extracts was evaluated under tensile load by stress—strain mechanical test. The stress—strain curves show similar behavior independent of the presence of cork extract and irganox as additives, or for neat PP. During the assay, it is observed that the break of the testing sample occurred at the maximum load or shortly after this. Therefore, for clarity and uniformity of results, it is presented the maximum strain at the yield point, for all specimens and for all conditions. The values obtained for tensile modulus, maximum strength and maximum strain at yield point are presented in Fig. 6.

The tensile modulus, maximum strength and maximum strain, measured for the processed mixtures, are statistically similar to the neat PP. Therefore, and despite the increase of the OOT and OIT, the results for the mechanical properties do not allow concluding about the protective effect of the extracts, during the processing stage (which occurs under poor oxygen atmosphere). Nevertheless, the mechanical evaluation indicates that the presence of the cork extracts does not negatively affect the mechanical performance of the PP matrix. The results presented further confirm the potential of the cork extracts to be explored as stabilizing and anti-oxidant additives to PP in a natural and eco-friendly perspective.

4. Conclusions

The presented results show that PP–cork composites have intrinsic protection against thermo-oxidation processes and that this protection is provided by cork, more specifically, by the ethanol soluble fraction present in the cork extractives fraction. After isolation of these components and its direct compounding with PP by twin-screw extrusion, it was demonstrated that the protective effect is retained. Mechanical analysis of the PP loaded with the cork extracts up to 1.5 wt.% show no compromise on the mechanical properties under tensile load. The feasibility of the cork extractives as bio-based thermal-oxidation stabilizers for PP has been demonstrated, opening new routes of valorization for this natural resource.

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